

Exploration of the diversity of Eukaryotic, Prokaryotic and Archaea communities in deep subsurface microbial ecosystems

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♦ Introduction

Exploration of microbial diversity within unique ecosystems worldwide contributes greatly to our understanding of the complexity and diversity of life. Such exploration has revealed the presence of numerous novel lineages of Bacteria, Archaea, and Eukaryota, many of which are uncultured; has expanded our knowledge of the limits of life; and has provided insight to physiological and biogeochemical processes. Here we build on previous studies exploring the diversity of the deepest levels accessible within the Sanford Underground Research Facility (SURF). The characterization of these microbial communities and fungal bodies will allow us to have better insight to the diversity of microbes that have come to inhabit areas of this subsurface environment, and provide clues to the nature of life when it is shielded from cosmic radiation, and survives in carbon-poor extreme environments.

- Earlier studies of Bacteria and Archaea diversity present in samples of soil, water and soil-like biofilms sampled from SURF early upon re-entry indicated the presence of novel taxa^{1, 2, 3}.
- In 2011, geoscientists Borgonie and Onstott discovered the deepest living animal, Halicephalobus mephisto in a south African gold mine at depths ranging from 4224-11616 ft underground⁴.

♦ Methods & Materials

ID of Fungi by sequencing of ITS region of rDNA:

- Fungal isolates were collected from various locations throughout the 4850 level at SURF and cultured in potato dextrose agar.
- PCR was used to amplify DNA with ITS1 and ITS4 primers using a Phire Plant Direct PCR Kit (ThermoFischer).
- The Applied Biosystems 3130xl genetic analyzer (Life Technologies) was used to perform Sanger sequencing on the amplified fungal DNA.
- The sequences were analyzed through Sequencher 5.2 (GeneCodes) and characterized using NCBI's nBLAST (nucleotide Basic Local Alignment Search Tool).

NextGen Sequencing of rDNA from Eukarya, Bacteria and Archaea within the Biofilms:

- Biofilms were collected from 4850 level of SURF. (Figure 1)
- DNA was extracted from the samples with the PowerLyzer PowerSoil **DNA** Isolation Kit.
- DNA was concentrated using a Zymo DNA Clean & Concentrator Kit.
- The 16S (Bacteria & Archaea) and 18S (Eukarya) rDNA from each biofilm was PCR amplified using Domain specific primers.
- The Metagenomic16S and 18S rDNA amplicon libraries were prepared for sequencing using the standard 16S Metagenomic Sequencing Preparation for the Illumina MiSeq system.
- Bacteria and Archaea 16S sequences were analyzed using the 16S analysis pipeline available in Illumina's BaseSpace System.
- Eukaryotic 18S sequences were analyzed through CLC Bio (Qiagen) for trimming and OTU clustering.
- nBLAST against GenBank (NCBI) was used to obtain the closest taxonomic matches for preliminary identification of diversity for each

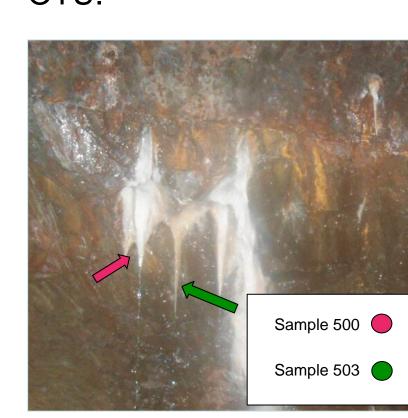


Figure 1. Two biofilms of a snottite-like texture differing in color were collected from a drift wall approximately 1 m from the drift floor. 500 appeared to be a light colored opaque white, while 503 had a brownish tinge. The biofilms were growing in an area where water was flowing from a drill hole that bathed these biofilms.

♦ Methods & Materials

Visual characterization:

- Regular light microscopy was used to scan small portions of the biofilm.
- Small portions of the biofilms were preserved for scanning electron microscopy (SEM) with a phosphate buffer containing 5% glutaraldehyde.
- The buffer was replaced by a series of ethanol washes with increasing concentrations until 100% has been reached.
- The sample was then placed under critical point drying followed by coating with a film of gold before placed under a scanning electron microscope.

♦ Results/Discussion

Fungi:

Amongst the variety of genera detected, Arthrobotrys stood out as the most interesting. Arthrobotrys is a carnivorous fungi that feeds on nematodes by entrapping the roundworm with it's hyphae. (Figure 2)

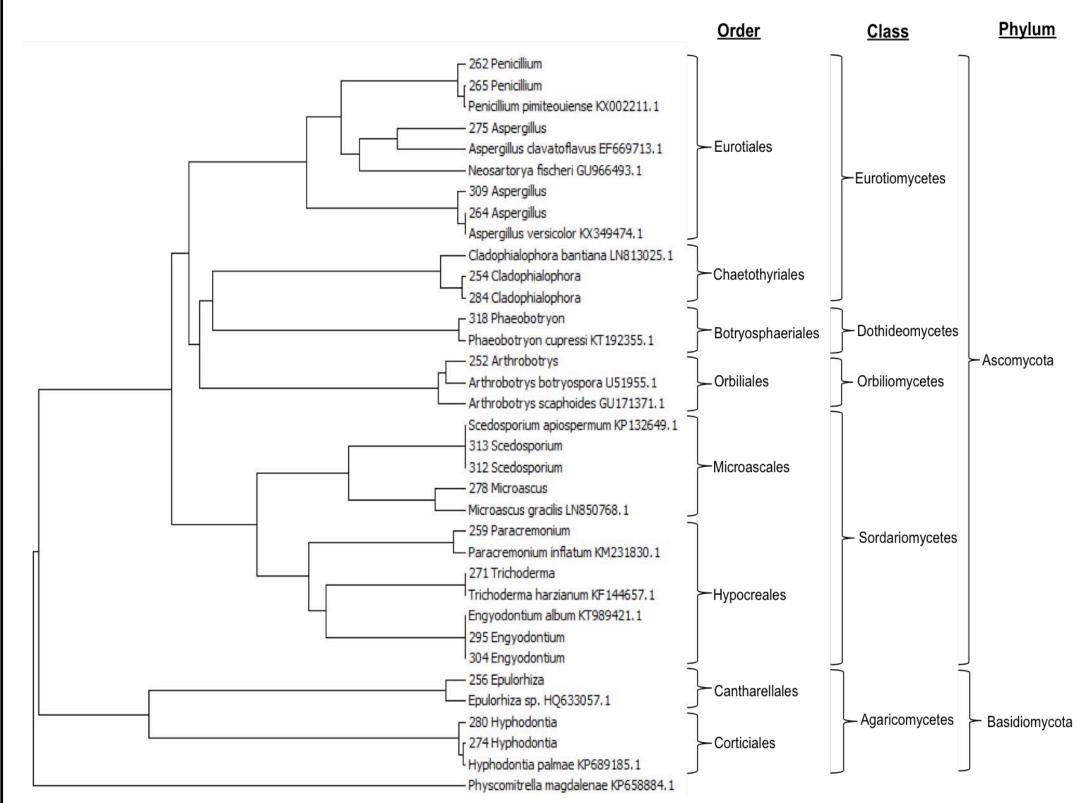
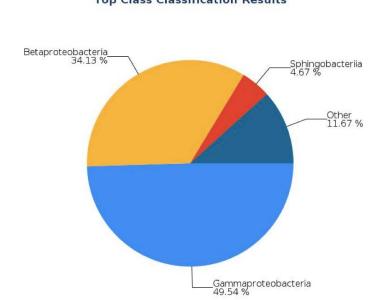


Figure 2 Phylogenic tree of fungal ITS sequences with reference sequences in GenBank. Phycomitrella magdalenae was selected as out-group to root the tree.

Bacterial life:

Sample 500

16S Metagenomics report indicates that the collected biofilms displayed a high abundance of Betaproteobacteria, Gammaproteobacteria and Bacteroidetes (Figure 3).



| | Thiobacillus thioparus | 2. |
|--|---|----|
| | Methyloversatilis universalis | 2. |
| Figure 3 (Above) Pie chart of class abundance within sample 500. | Figure 4 (Above) Table representing the top 5 species | |

Methylocaldum tepidum

Methylobacillus glycogenes

% Total Reads

- present in sample 500. Amongst the species found with the highest abundance, it is possible that Methylocaldum tepidum (Gammaproteobacteria), a methane-oxidizing bacteria³ and Methylobacillus glycogenes (Betaproteobacteria), a glycogen-producing bacteria utilize one another's metabolites. (Figure 4).
- Comparing previous studies performed at the Homestake mine, our results reveal some consistency with the most abundant Phylum identified in the soil sample from the Ross shaft wall studied by Waddell et al, 2010, but it differs significantly in abundance at the species level. The soil sample had a high abundance of aerobic sulfur-oxidizing bacteria of the *Thiobacillus*¹.

♦ Results/Discussion

Bacterial life: Sample 503

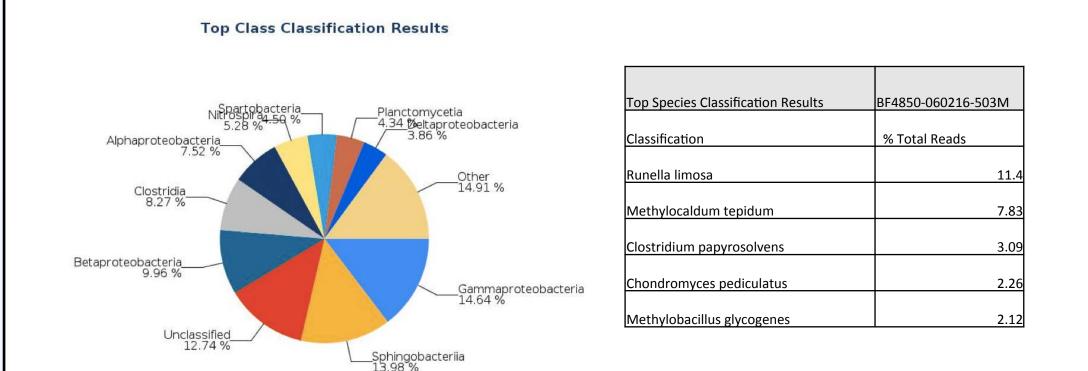


Figure 5 (Above) Pie chart of class abundance within sample 503.

Figure 6 (Above) Table representing the top 5 species present in sample 500.

503 exhibited a much more diverse variety of bacterial classes with Gammaproteobacteria and Sphingobactera being the most abundant.

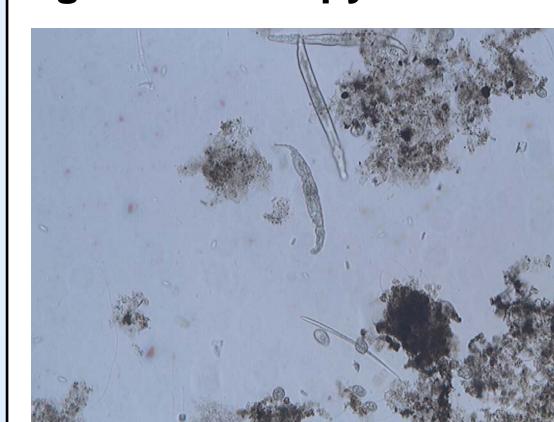
Archaea life:

In contrast to the diversity found in the bacterial results, over 90% of the Archaea data were found to belong to the phylum Thaumarchaeota and grouped most closely to Nitrosopumilus maritimus, a common ammonia oxidizer that is usually found in non-coastal marine environments⁵.

Eukayotic life:

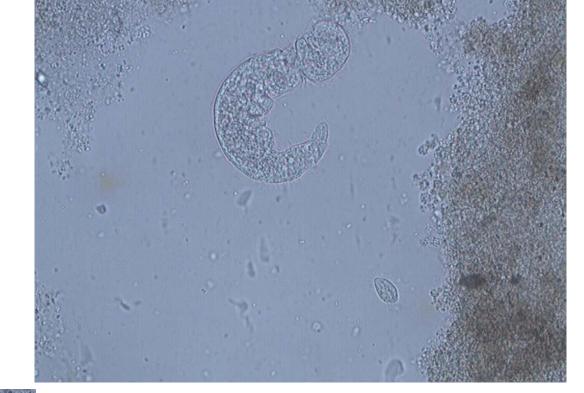
Contrary to common belief, the underground is not the desolate abyss that we imagine it to be, but rather a world blooming with microscopic life and communities that await further discovery.

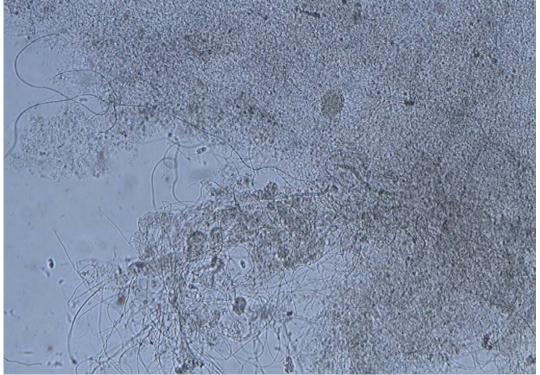
Light Microscopy



Sample 503 under Light Microscopy at 100X(Left) Multiple unidentified bilaterian and nematode.

Sample 500 under light microscopy at 400X (Right) Unidentified Eukaryotes.





100X (Left) Filamentous biofilm with a variety of microbes.

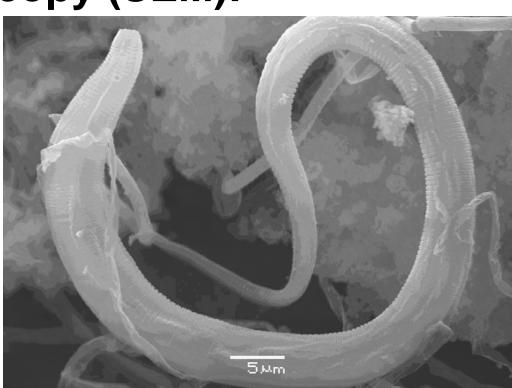
Sample 500 under light microscopy at

Unidentified flagellate by light microscopy at 400X (Left) uses its single flagellum to propel itself forward and obtain sensory information.

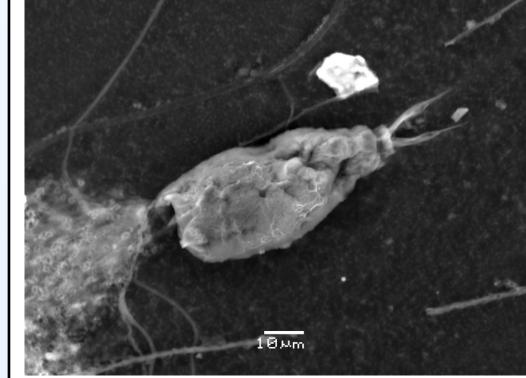


Scanning Electron Microscopy (SEM):

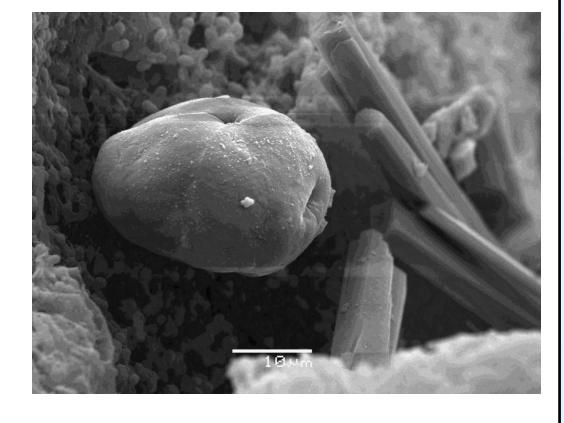
Nematode under SEM (Right) presence supported by NextGen Metagenomic data that still awaits further analysis for specific genus.



Rotifer under SEM (Left)



Suspected pollen granule under SEM (Right) 2500X



♦ References

- 1. Waddell, E.J., Elliott, T.J., Sani, R.K., Vahrenkamp, J.M., Roggenthen, W.M., Anderson, C.M., & Bang, S.S. (2010). Phylogenetic evidence of noteworthy microflora from the subsurface of the former Homestake gold mine, Lead, South Dakota. Environmental Technology 31(8-9):979–991 http://doi.org/10.1080/09593331003789511Waddell, E.J. et al. 2010. Environmental Technology 31:997-991
- 2. Moser, D.P., Anderson, C., Jones, T., Bang, S., Kieft, T., Boutt, D., Sherwood-Lollar, B., Onstott, T.C., Murdoch, L., Pfiffner, S., Bruckner, J., Fisher, J., Newburn, J. (2010). First Microbial Community Assessment of Borehole Fluids from the Sanford Lab/Deep Underground Science and Engineering Laboratory (DUSEL). Abstract H13F-1049 presented at 2010 Fall Meeting, AGU, San Francisco, Calif., 13-17 Dec.
- Rastogi, R., Stetler, L.D., Peyton, B.M. and Sani, R.K. (2009). Molecular Analysis of Prokaryotic Diversity in the Deep Subsurface of the Former Homestake Gold Mine, South Dakota, USA. The Journal of Microbiology 47(4):371-384
- 4. Borgonie, G., Linage-Alvarez, B., Ojo, A. O., Mundle, S. O. C., Freese, L. B., Van Rooyen, C., Kuloyo, O., Albertyn, J., Pohl, C., Cason, E. D., Vermueulen, J., Pienaar, C., Litthauer, D., Van Niekerk, H., Van Eeden, J., Sherwood-Lollar, B., Onstott, T. C., Van Heerden, E. (2015). Eukaryotic opportunists dominate the deep-subsurface biosphere in South Africa. *Nature* Communications 6:8952.
- Takeuchi M, Kamagata Y, Oshima K, Hanada S, Tamaki H, Marumo K, Maeda H, Nedachi M, Hattori M, Iwasaki W, Sakata S. (2014) Int J Syst Evol Microbiol. Sep;64(Pt 9):3240-6. doi: 10.1099/ijs.0.063503-0. Epub 2014 Jun 30.

♦ Acknowledgements

This project was funded by NSF REU grant #1560474. I would like to thank Tom Regan, Dr. Brianna Mount and others in Sanford Underground Research Facility who made this possible.

Special thanks to WestCORE, specifically Oxana Gorbatenko and Bethany Reman for their time and efforts invested into this project.

Gratitude to my mentor, Dr. Cynthia Anderson for her knowledge, patience and guidance.



